

**ANNEX 1**

Component (reference)	Analysis			
	Mode of isolation	Réf.	Identification/Quantification	Ref.
<b>Amino compounds</b>				
alanine; (8,16,23,28,45,46,47,48,50)	IE, RE	16,53	AAA, GCMS, SP	16,23,34,53,54,56
asparagine; (8,16,23,28,46,50)	IE	16	AAA, SP	16,23,28
leucine/isoleucine; (8,16,23,28,46,47,48,50)	IE, RE	16,47	AAA, SP, GCMS	16,23,28,47,48,50
valine; (8,16,23,28,46,47,50)	IE, RE	16	AAA, SP, GC	16,23,28,50
glutamine; (8,16,23,28,46,50)	IE	16	AAA, SP	16,23,28
serine/homoserine; (8,16,23,28,46,47,50)	IE, RE	16,47	AAA, SP, GC	16,23,28,47,50
glycine; (8,16,23,22,28,46,47,50)	IE, RE	16,47	AAA, SP, GC	16,23,28,47,50
phenylalanine; (8,16,18,23,46,47,48)	IE, RE	16,47	HPLC, AAA, GC	16,18,23,47,50
threonine; (8,16,23,46,47)	IE, RE	16,47	AAA	16,23,47
tyrosine; (8,16,18,26,46,47,50)	IE, RE	16,47	HPLC, AAA, GC	16,18,26,47,50
lysine; (8,16,23,22,28,46,47,50)	IE, RE	16,47	AAA, SP, GC	16,23,28,47,50
proline; (8,16,23,46,50)	IE	16	AAA, GC	16,23,50
methionine; (8,16,23,46,50)	IE	16	AAA, GC	16,23,50
cystathionine; (8,46)				
ornithine; (8,16,23,46,50)	IE	16	AAA, GC	16,23,50
citrulline; (23,16)	IE	16	AAA	16,23
arginine; (8,11,16,28,46)	IE	16	AAA, SP	16,23,28
glutamate; (47,48,50)	RE, IE	47	GCMS, AAA	47,48,50
aspartate; (47,48,23,50)	RE, IE	47	GCMS, AAA	47,48,50
tryptophan; (8,18)			HPLC	18
histidine; (8,23,16,46,47)	IE, RE	16,47	AAA	16,23,47
cysteic acid; (8,46)				
aspartic acid; (8,16,23,28,46)	IE	16	AAA, SP	16,28
glutamic acid; (8,16,23,22,28,46)	IE	16	AAA, SP	16,23,28
-amino butyric acid; (8,16,28,46)	IE	16	AAA, SP	16,28
amino adipic acid; (16)	IE	16	AAA	16
ethanolamine; (16)	IE	16	AAA	16
2,4-dihydroxy-1,4-benzoxazin-3-one; (34)	XAD-4	34	HPLC, GC	34
ammonium; (37)			Biotronic	37
ammonia; (8)				
cystine; (16,46)	IE	16	AAA	16
benzoxazolin-2-one; (34)	XAD-4	34	HPLC, GC	34
6-methoxybenzolin-2-one; (34)	XAD-4	34	HPLC, GC	34
2,4-dihydroxy-7-methoxy-1,4-benzoxazin-3-one; (34)	XAD-4	34	HPLC, GC	34
<b>Organic acids</b>				
oxalic acid; (8,37,46,47)	RE, IE	47	UV/Vis, HPLC	37,47
malic acid; (8,21,22,24,25,28,28,30,37,46,47)	RE, IE	30,47	UV/Vis, GC HPLC, IC, MS	24,28,28,30,37,47
acetic acid; (8,46)				
propionic acid; (8,46)				
butyric acid; (8,46)				
valeric acid; (8,46)				
citric acid; (8,11,21,22,24,25,26,28,28,30,37,46,47)	RE, IE	30,47	QEA, Xspec, UV/Vis, IC, HPLC, GC, MS	8,24,28,28,30,37,47
succinic acid; (8,24,28,28,30,37,46,47)	RE, IE	28,30,47	UV/Vis, GC HPLC, IC, MS	24,28,28,30,37,47

fumaric acid; (8,28,29,37,47)	RE, IE	47	UV/Vis, GC HPLC, MS	28,29,37,47
glycolic acid; (8,46)				
deoxymugineic acid; (1)				
malonic acid; (8)				
2-ketogluconic acid; (38)	IE	38	GC, TLC, Xdif, SR, AA	38
tartaric acid; (8,29,37,47)	RE,IE	47	UV/Vis, GC, HPLC	29,37,47
isocitric acid; (37)			HPLC,UV/Vis	37
aconitic acid; (29,47)	RE, IE	47	UV/Vis, HPLC	47
3-phenyl propionic acid; (56)	XAD-4	56	GCMS	56
p-hydroxybenzoic acid; (4,9,41,54,56)	XAD-4	41,56	HPLC,GCMS	4,41,56
2,5-dihydroxybenzoic acid; (56)	XAD-4	56	GCMS	56
myristic acid; (56)	XAD-4	56	GCMS	56
p-hydroxycinnamic acid; (52,56)	XAD-4	56	GCMS	56
palmitic acid; (8,56)	XAD-4	56	GCMS	56
aconitic acid; (29)			GC	29
stearic acid; (8,56)	XAD-4	56	GCMS	56
oxalocetic acid; (29)	concentration	29	GC	29
uronic acid; (38)				
glutaric acid; (29)	concentration	29	GC	29
glyoxylic acid; (29)	concentration	29	GC	29
pentadecanoic acid; (52)	XAD-4	52	GCMS	52
<b>Carbohydrates</b>				
glucose; (8,16,29,38,46,47,48,50)	IE, RE, MF	47,50	GCMS, HPLC UV/Vis	16,29,47,48,50
fructose; (8,16,29,38,46,47,48,50)	IE, RE, MF	47,50	GCMS, HPLC UV/Vis	16,29,47,48,50
maltose; (8,46)				
galactose; (8,46,47)	IE, RE	47	UV/Vis, HPLC	47
ribose; (8,46,47,48)	IE, RE	47	GCMS, HPLC UV/Vis	47,48
xylose; (8,38,46,47)	IE, RE	47	UV/Vis, HPLC	47
rhamnose; (8,46)				
arabinose; (8,29,46,47)	IE, RE	47	UV/Vis, GC, HPLC	29,47
raffinose; (8,46)				
oligosaccharides; (8,46)				
myo-inositol; (50)	MF	50	GCMS, HPLC	50
deoxyribose; (8)				
sucrose; (8,16,29,47,48,50)	IE, RE, MF	47,50	GCMS, HPLC UV/Vis	16,29,47,48,50
deoxysugars; (8)				
<b>Phenolic compounds</b>				
salicylic acid; (54)	XAD-4	54	MNR, SP	54
p-hydroxybenzoic acid; (4,9,41,54)	XAD-4	41,54	MNR, SP, HPLC	4,41,54
vanillic acid; (4,41,54)	XAD-4	41,54	HPLC, MNR, SP	4,41,54
syringic acid; (4,15,52,54)	XAD-4, XAD-2	15,52,54	GCMS, MNR, SP, HPLC	4,15,52,54
4-methoxyindole-3-acetonitrile; (54)	XAD-4	54	MNR, SP	54
pyrocatechol; (54)	XAD-4	54	MNR, SP	54
coumesterol; (9, 43,44)			HPLC	18
caffeic acid; (18,26)			HPLC	18
p-thiocyanatophenol; (56)	XAD-4	56	GCMS	56
2-hydroxybenzothiazole; (56)	XAD-4	56	GCMS	56
3,4-dimethylbenzoic acid; (52)	XAD-4	52	GCMS	52
benzoic acid; (18,29,52,56)	XAD-4	52,56	HPLC, GC, MS	18,29,52,56
phenylacetic acid; (52)	XAD-4	52	GCMS	52
2-methoxyphenol; (52)	XAD-4	52	GCMS	52
hydrocinnamic acid; (52)	XAD-4	52	GCMS	52
cinnamic acid; (18,52,56)	XAD-4	52,56	HPLC, GCMS	18,52,56
2-methoxy phenylacetic acid; (52)	XAD-4	52	GCMS	52
3-hydroxy hydrocinnamic acid; (52)	XAD-4	52	GCMS	52

4-hydroxy-3-methoxy hydrocinnamic acid; (52)	XAD-4	52	GCMS	52
4-hydroxy-2-methoxycinnamic acid; (52)	XAD-4	52	GCMS	52
ferulic acid; (4,6,14,18,52)	XAD-4	52	HPLC, GCMS	4,18,52
cyclopropyl-p-benzoquinone (14)				
2,6-dimethoxy-p-benzoquinone (14)				
tetrafluorbenzoquinone (14)				
benzoquinone (14)				
SXSg (14)				
strigol (14)				
resorcinol (14)				
dihydroquinone (14)				
sinapic acid; (15,52)	XAD-4, XAD-2	15,52	GCMS, HPLC	15,52
2-(3',5'-dihydroxyphenyl)-5,6-dihydroxy-benzofuran; (34)	XAD-4	34	HPLC, MS, NMR, UV/Vis	34
<b>Flavonoids</b>				
kievitone; (26)			HPLC	26
4',7-dihydroxyflavone; (9,12,15,19,35,36,44)	XAD-2, CF, HPLC	9,15,36	HPLC, MS, NMR, EP, UV/Vis	9,12,15,35,44
4',7-dihydroxyflavanone; (9,12,15,19,35,36,44)	CF, HPLC	36	HPLC, MS, NMR, UV/Vis	35,36
formononetin-4',7-dihydroxyflavonone; (9,19,35,36,44)	CF, HPLC	36	HPLC, MS, NMR, UV/Vis	35,36
4',5,7-dihydroxyflavonone; [apigenin] (9,18,26,43)			HPLC	18,26
apigen-7-O-glucoside; (9,15)	XAD-2	15	MS, HPLC, EP	15
genistein; (15,17,18,43)	XAD-2	15	HPLC, MS, EP	15,18
3',4',5,7-tetrahydroxyflavone; [leuteolin] (9,15,18,15,26,42,43)	XAD-2	15	HPLC, EP, MS NMR, UV/Vis	15,18,26,42
4',7-dihydroxyisoflavone; [daidzein] (9,15,17,18,43,44)	XAD-2, CF, HPLC	9,15,17,44	EP, HPLC, MS, UV/Vis, NMR	9,15,17,18,44
3,4',5,7-tetrahydroxy flavone; [kaempferol] (9,15,18,26,43)	XAD-2	15	HPLC, EP, MS NMR, UV/Vis	15,18,26,43
coumestrol; (9,43,44)	HPLC	9	HPLC, UV/Vis	9
formononetin-7-O-(6"-O-malonylglucoside); (9,10)	CF, HPLC	10	MS, NMR, UV/Vis	10
formononetin; (9,14,18,36,44)	CF, HPLC	36,44	HPLC, NMR, MS, UV/Vis	9,18,36,44
3',4',7-trihydroxyflavone; (9,15)	HPLC	9	UV/Vis	9
4',7-dihydroxy-3-methoxyflavone; [geraldone] (9,12,44)	HPLC	9,44	HPLC, NMR, UV/Vis	9,12,44
4'-hydroxy-7-methoxyflavone; (9,44)	HPLC	44	HPLC, NMR	9,44
xenognosin A & B (14)				
<b>Enzymes, Nucleotides &amp; Chalcones</b>				
invertase; (46,8)				
amylase; (46,8)				
protease; (46,8)				
guanine; (46,8)				
adenine; (46,8)				
polygalacturonase; (8)				
phosphatase; (7,8)				7
uridine/cytidine; (8)				
4,4'-dihydroxy-2'-methoxychalcone; (10,19,35,36)	CF, HPLC	36	HPLC, MS, NMR, UV/Vis	10,35,36
<b>Fatty acids and stérols</b>				
cholesterol; (8)				
palmitic acid; (8)				
-sitosterol; (8,50)	EP, TLC	50	GCMS	50
stigmasterol; (8,50)	EP, TLC	50	GCMS	50
campesterol; (50,8)	EP, TLC	50	GCMS	50
stearic acid; (8)				

oleic acid; (8)				
linoleic acid; (8)				
Acides gras 18:1; 18:2; 18:3; 20:0; 22:0; 24:0; (50)	chromatography	50	GCMS, HPLC	50
<b>Others</b>				
epi-3-hydroxy-mugineic acid; (2,45,53)	HPLC	53	HPLC	53
8-methylsulfinyloctyl isothiocyanate [histurin]; (54)	XAD-4	54	NMR, SP	54
benzyl isothiocyanate; (51,54)	XAD-4	51	GC	51
auxins; (8,32)				
scopoletin; (8,41)	XAD-4	41	HPLC	41
fluorescent substances; (8)				
vitamins; (8)				
hydrocyanic acid; (8)				
glycosides; (8)				
saponines; (8)				
Composés organiques phosphorés; (8)				
nematode cyst or egg hatching factors; (8,46)				
nematode attractants/nematocides; (8,46)				
fungal mycelium stimulants and inhibitors; (5,8,13)				
zoospore attractants; (5,8,33,46)				
spore and aclerotium germination stimulants and inhibitors; (5,8,39)				
parasitic weed germination stimulants; (8,39)	XAD-4	39	HPLC	39
medicarpins; (8,10,34)	CF, HPLC	10	MS, NMR, UV/Vis	10
medicarpin-3-O-glycoside; (8,10)	CF, HPLC	10	MS, NMR, UV/Vis	10
umbelliferone; (9,43,44)			HPLC, NMR	9,44
coumarins; (4,9,41,43)	XAD-4	41	HPLC	4,41
modulation gene inducers; (8,43)				
assorted allelopathic compounds; (6,8,55)	XAD-4	55		
metal chelators; (8)				
ethanol; (47)			GC	48
methanol; (8)				
formaldehyde; (8)				
acetaldehyde; (8,48)				
proionaldehyde; (8)				
acetone; (8)				
ethylene; (8)				
propylene; (8)				
various volatiles; (3,5)				
gibberellins; (8, 18)			HPLC	18
cytokinins; (8)				

IE=ion exchange trap; GC=gas chromatography; HPLC=high performance liquid chromatography; MS=mass spectrometry; RE=rinse & evaporation; AAA=automatic amino-acid analyzer; NMR=nuclear magnetic resonance, CF=centrifugation; EP=electrophoresis; SP=spectrophotometry; MF=membrane filtering; TLC=thin layer chromatography; Xdiff= X-ray diffraction

## References

- 1) Award F, Römheld V, Marschner H (1994) Effect of root exudates on mobilization in the rhizosphere and uptake of iron by wheat. Plant and Soil 165:213-218
- 2) Bar-Ness E, Hadar Y, Chen Y, Romhfeld V, Marschner H. (1992) Short-term effects of rhizosphere microorganisms on Fe uptake from microbial siderophores by maize and oat. Plant Physiol. 100:451-456
- 3) Bécard G, Piché Y (1989) Fungal growth stimulation by CO<sub>2</sub> and root exudates in vesicular-arbuscular mycorrhizal symbiosis. Appl. Environ. Microbiol. 55(9):2320-2325
- 4) Ben-Hammouda M, Kremer RJ, Minor HC, Sarwar M. (1995) A chemical basis for differential allelopathic potential of sorghum hybrids on wheat. J. Chem. Ecol. 21(6):775-786
- 5) Benizri E, Courtade A, Guckert A (1995) Fate of two microorganisms in maize simulated rhizosphere under hydroponic and sterile conditions. Soil Biol. Biochem. 27(1):71-77

- 6) Blum U, Dalton BR (1985) Effects of ferulic acid, an allelopathic compound, on leaf expansion of cucumber seedlings grown in nutrient culture. *J Chem. Ecol.* 11(3):279-301
- 7) Cumming JR, Weinstein LH (1990) Utilization of  $\text{AlPO}_4$  as a phosphorous source by ectomycorrhizal *Pinus rigida* Mill. seedlings. *New Phytol.* 116:99-106
- 8) Curl EA, Truelove B (1986) *The Rhizosphere*. Springer-Verlag, Berlin. pp288
- 9) d'Arcy-Lameta A (1986) Study of soybean and lentil root exudates II. Identification of some polyphenolic compounds, relation with plantlet physiology. *Plant and Soil* 92:113-123
- 10) Dakora FD, Joseph CM, Phillips DA (1993) Alfalfa (*Medicago sativa* L.) root exudates contain isoflavonoids in the presence of *Rhizobium meliloti*. *Plant Physiol.* 101:819-824
- 11) Dinkelaker B, Römheld V, Marschner H (1989) Citric acid excretion and precipitation of calcium citrate in the rhizosphere of white lupin (*Lupinus albus* L.). *Plant Cell Environ.* 12:285-292
- 12) Djordjevic MA, Redmond JW, Batley M, Rolfe BG (1987) Clovers secrete specific phenolic compounds which either stimulate or repress nod gene expression in *Rhizobium trifolii*. *EMBO J.* 6(5):1173-1179
- 13) Elias KS, Safir GR (1987) Hyphal elongation of *Glomus fasciculatus* in response to root exudates. *Appl. Environ. Microbiol.* 53:1928-1933
- 14) Estabrook EM, Yoder JL (1998) Plant-plant communications: rhizosphere signaling between parasitic angiosperms and their hosts. *Plant Physiol.* 116:1-7
- 15) Firmin JL, Wilson KE, Rossen L, Johnston AWB (1986) Flavonoid activation of nodulation genes in *Rhizobium* reversed by other compounds present in plants. *Nature* 324:90-92
- 16) Gamliel A, Katan J (1992) Influence of seed and root exudates on fluorescent pseudomonads and fungi in solarized soil. *Phytopathology* 82:320-327
- 17) Graham TL (1991) Flavonoid and isoflavonoid distribution in developing soybean seedling tissue and in seed and root exudates. *Plant Physiol.* 95:594-603
- 18) Graham TL (1991) A rapid, high resolution high performance liquid chromatography profiling procedure for plant and microbial aromatic secondary metabolites. *Plant Physiol.* 95:584-593
- 19) Hartwig UA, Maxwell CA, Joseph CM, Phillips DA (1989) Interactions among flavonoid nod gene inducers released from alfalfa seeds and roots. *Plant Physiol.* 91:1138-1142
- 20) Hausenbuiller RL (1985) *Soil Science: Principles & Practices 3<sup>rd</sup> Edition*. Wm. C. Brown Company Publisher, Dubuque, Iowa. p. 378-381
- 21) Hoffland E (1992) Quantitative evaluation of the role of organic acid exudates in the mobilization of rock phosphate by rape. *Plant and Soil* 140:279-289
- 22) Hoffland E, Findenegg GR, Nelemans JA (1989) Solubilization of rock phosphate by rape II. Local root exudation of organic acids as a response to P-starvation. *Plant and Soil* 113:161-165
- 23) Ivarson KC, Sowden FJ, Mack AR (1970) Amino-acid composition of rhizosphere as affected by soil temperature, fertility and growth stage. *Can. J. Soil sci.* 50:183-189
- 24) Johnson JF, Allan DL, Vance CP, Weiblen G (1996) Root carbon dioxide fixation by phosphorus-deficient *Lupinus albus*: contribution to organic acid exudation by proteoid roots. *Plant Physiol.* 112:19-30
- 25) Jones DL, Darrah PR (1994) Role of root derived organic acids in the mobilization of nutrients from the rhizosphere. *Plant and Soil* 166:247-257
- 26) Jones DL, Edwards AC, Donachie K, Darrah PR (1994) Role of proteinaceous amino acids released in root exudates in nutrient acquisition from the rhizosphere. *Plant and Soil* 158:183-192
- Kapulnik Y, Joseph CM, Phillips DA (1987) Flavone limitations to root nodulation and symbiotic nitrogen fixation in alfalfa. *Plant Physiol.* 84:1193-1196
- 27) Kloss M, Iwannek KH, Fendrik I, Niemann EG (1984) Organic acids in the root exudates of *Diplachne fusca* (Linn.) Beauv. *Environmental and Experimental Botany* 24(2):179-188
- 28) Krafczyk I, Trolldenier G, Beringer H (1984) Soluble root exudates of maize: influence of potassium supply and rhizosphere microorganisms. *Soil Biol. Biochem.* 16(4):315-322
- 29) Lipton DS, Blanchar RW, Blevins DG (1987) Citrate, malate, and succinate concentration in exudates from P-sufficient and P-stressed *Medicago sativa* L. seedlings. *Plant Physiol.* 85:315-317
- 30) Lynch JM (1990) *The Rhizosphere*. John Wiley & Sons, Chichester, England. pp. 458.
- 31) Martinez-Toledo MV, de la Rubia T, Moreno J, Gonzalez-Lopez J (1988) Root exudates of *Zea mays* and production of auxins, gibberellins and cytokinins by *Azotobacter chroococcum*. *Plant and Soil* 110:149-152
- 32) Martinez-Toledo MV, Moreno J, de la Rubia T, Gonzalez-Lopez J (1988) Root exudates of *Zea mays* and the acetylene-reduction activity of *azotobacter chroococcum*. *Soil Biol. Biochem.* 20(6):961-962
- 33) Masaoka Y, Kojima M, Sugihara S, Yoshihara T, Koshino M, Ichihara A (1993) Dissolution of ferric phosphate by alfalfa (*Medicago sativa* L.) root exudates. *Plant and Soil* 155/156:75-78
- 34) Maxwell CA, Hartwig UA, Joseph CM, Phillips DA (1989) A chalcone and two related flavonoids released from alfalfa roots induce nod genes of *Rhizobium meliloti*. *Plant Physiol.* 91:842-847
- 35) Maxwell CA, Phillips DA (1990) Concurrent synthesis and release of nod-gene-inducing flavonoids from alfalfa roots. *Plant Physiol.* 93:1552-1558
- 36) Mench M, Martin E (1991) Mobilization of cadmium and other metals from two soils by root exudates of *Zea mays* L., *Nicotiana tabacum* L. and *Nicotiana rustica* L. *Plant and Soil* 132:187-196
- 37) Moghimi A, Tate ME, Oades JM (1978) Characterization of rhizosphere products especially 2-ketogluconic acid. *Soil Biol. Biochem.* 10:283-287
- 38) Müller S, Van Der Merwe A, Schildknecht H, Visser JH (1993) An automated system for large-scale recovery of germination stimulants and other root exudates. *Weed Science* 41:138-143

- 39) Pérez FJ, Ormeño-Núñez J (1991) Root exudates of wild oats: allelopathic effect on spring wheat. *Phytochemistry* 30(7):2199-2202
- 40) Pérez FJ, Ormeño-Núñez J (1991) Difference in hydroxamic acid content in roots and root exudates of wheat (*Triticum aestivum* L.) and rye (*Secale cereale* L.): possible role in allelopathy. *J. Chem. Ecol.* 17(6):1037-1043
- 41) Peters NK, Frost JW, Long SR (1986) A plant flavone, luteolin, induces expression of *Rhizobium meliloti* nodulation genes. *Science* 233:977-979
- 42) Peters NK, Long SR (1988) Alfalfa root exudates and compounds which promote or inhibit induction of *Rhizobium meliloti* nodulation genes. *Plant Physiol.* 88:396-400
- 43) Redmond JW, Batley M, Djordjevic MA, Innes RW, Kuempel PL, Rolfe BG (1986) Flavones induce expression of nodulation genes in *Rhizobium*. *Nature* 323(16):632-634
- 44) Römhild V (1991) The role of phytosiderophores in acquisition of iron and other micronutrients in graminaceous species: an ecological approach. *Plant and Soil* 130:127-134
- 45) Rovira AD (1969) Plant root exudates. *Bot. Rev.* 35:35-57
- 46) Schwab SM, Leonard RT, Menge JA (1984) Quantitative and qualitative comparison of root exudates of mycorrhizal and nonmycorrhizal plant species. *Can. J. Bot.* 62:1227-1231
- 47) Smucker AJM, Erickson AE (1987) Anaerobic stimulation of root exudates and disease of peas. *Plant and Soil* 99:423-433
- 48) Son K, Severson RF, Snook ME, Kays SJ. (1991) Root carbohydrate, organic acids, and phenolic chemistry in relation to sweetpotato weevil resistance. *HortScience* 26(10):1305-1308
- 49) Svenningsson H, Sundin P, Liljenberg C (1990) Lipids, carbohydrates and amino acids exuded from the axenic roots of rape seedlings exposed to water-deficit stress. *Plant Cell Environ.* 13:155-162
- 50) Tang CS, Takenaka T (1983) Quantitation of a bioactive metabolite in undisturbed rhizosphere-benzyl isothiocyanate from *Carica papaya* L. *J. Chem. Ecol.* 9(8):1247-1253
- 51) Tang CS, Young CC (1982) Collection and identification of allelopathic compounds from the undisturbed root system of bigalta limpgrass (*Hemarthra altissima*). *Plant Physiol.* 69:155-160
- 52) Treeby M, Marschner H, Römhild V (1989) Mobilization of iron and other micronutrient cations from a calcareous soil by plant-borne, microbial, and synthetic metal chelators. *Plant and Soil* 114:217-226
- 53) Yamane A, Nishimura H, Mizutani J (1992) Allelopathy of yellow fieldcress (*Rorippa sylvestris*): identification and characterization of phytotoxic constituents. *J. Chem. Ecol.* 18(5):683-691
- 54) Young CC (1984) Autointoxication in root exudates of *Asparagus officinalis* L. *Plant and Soil* 82:247-253
- 55) Yu JQ, Matsui Y (1994) Phytotoxic substances in root exudates of cucumber (*Cucumis sativus* L.). *J. Chem. Ecol.* 20(1):21-31